
PART I - ADMINISTRATIVE

Section 1. General administrative information

Title of project

Evaluate Effects Of Catch And Release Angling On White Sturgeon

BPA project number: 20063

Contract renewal date (mm/yyyy): ☐ Multiple actions?

Business name of agency, institution or organization requesting funding

U.S. Geological Survey, Columbia River Research Laboratory, Idaho Department of Fish and Game

Business acronym (if appropriate) USGS, IDFG

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NPPC Program Measure Number(s) which this project addresses

10.1, 10.4A.4

FWS/NMFS Biological Opinion Number(s) which this project addresses

Other planning document references

Idaho Department of Fish and Game Fish Management Plan, 1996-2000, Columbia Basin Fish and Wildlife Authority Draft Multi-year Implementation Plan, Upper Snake River White Sturgeon Biological Risk Assessment

Short description

Use physiological telemetry to monitor metabolic activity, determine energetic costs and assess stressful effects of catch and release angling on white sturgeon.

Target species

White sturgeon (*Acipenser transmontanus*)

Section 2. Sorting and evaluation

Subbasin
Snake River

Evaluation Process Sort

CBFWA caucus	Special evaluation process	ISRP project type
Mark one or more caucus	If your project fits either of these processes, mark one or both	Mark one or more categories
<input type="checkbox"/> Anadromous fish <input checked="" type="checkbox"/> Resident fish <input type="checkbox"/> Wildlife	<input checked="" type="checkbox"/> Multi-year (milestone-based evaluation) <input type="checkbox"/> Watershed project evaluation	<input type="checkbox"/> Watershed councils/model watersheds <input type="checkbox"/> Information dissemination <input type="checkbox"/> Operation & maintenance <input type="checkbox"/> New construction <input checked="" type="checkbox"/> Research & monitoring <input type="checkbox"/> Implementation & management <input type="checkbox"/> Wildlife habitat acquisitions

Section 3. Relationships to other Bonneville projects

Umbrella / sub-proposal relationships. List umbrella project first.

Project #	Project title/description

Other dependent or critically-related projects

Project #	Project title/description	Nature of relationship
8605000	White Sturgeon Mitigation and Restoration in the Columbia and Snake Rivers	Equipment and expertise developed under 860500 project will be used in this proposed work. Fishery managers contemplating seasonal and area closures to angling will benefit from the proposed study.
9700900	Evaluate Means of Rebuilding White Sturgeon Population in Lower Snake River	Adaptive management to rebuild populations will benefit from knowing the effects of angling stress.

Section 4. Objectives, tasks and schedules

Past accomplishments

Year	Accomplishment	Met biological objectives?

Objectives and tasks

Obj 1,2,3	Objective	Task a,b,c	Task
1	Examine the relation between telemetered physiological variables, oxygen consumption, and physiological indicators of stress in white sturgeon.	a	Obtain or construct a swimming respirometer for large fish. Setup wet laboratory holding and testing facilities
		b	Obtain a few physiological telemetry tags and related equipment for lab testing. These would include heart rate, ventilation rate, and electromyogram (EMG) tags. Obtain detailed instructions and methods for surgical implantation of tags.
		c	Collect white sturgeon from the Columbia River and transport them to our laboratory. Maintain fish under ambient conditions. Fish should be about 0.9-1.5 m in length. Hold fish for several weeks prior to testing
		d	Implant one tag in each fish. Cannulate fish for repeated sampling of blood. Check for proper operation of the biotelemetry system. Allow for recovery while telemetering the variable of interest and maintaining patency of

			cannula.
		e	Subject fish to a respirometry trial. Conduct a U-critical swimming challenge with fish while simultaneously measuring oxygen consumption, monitoring telemetry signals (i.e., heart rate, ventilation rate, or EMG), and periodically taking blood samples
		f	Examine relations between telemetered data, oxygen consumption, physiological indicators of stress, and individual fish. Derive best fit regression equation for the relation between telemetered data and oxygen consumption.
2	Assess the effects of catch and release angling on metabolic rate and selected physiological indicators of stress in wild white sturgeon	a	From the wild, capture several large sturgeon with either set lines or angling. Implant tags and monitor recovery.
		b	Monitor physiological function and movement for several weeks to establish baseline metabolic rate data.
		c	Using the telemetry system, locate tagged fish and capture by angling. Monitor physiological function during the angling process. Monitor time of hookup and playing time. Upon landing the fish, draw a quick blood sample from the caudal vasculature.
		d	Determine the stressful effects of angling. After release, monitor physiological function for several days. Compare metabolic response of angling with the baseline metabolic rate data collected prior to angling.
3	Document the occurrence and	a	Use physiological telemetry and

	determine the extent of any postangling mortality		fish movement and location data to assess post-angling mortality.
4	Assess the effects of exhaustive stress on selected aspects of white sturgeon reproductive performance	a	Obtain several reproductive age female sturgeon from the wild. Stock fish in each of 4 large tanks for rearing. There will be two treatments (stressed and controls) with two replicate tanks per treatment. Maintain under ambient conditions.
		b	During rearing, monitor reproductive physiology and state of maturation by periodically sampling blood for sex steroid analysis and conducting ovarian biopsies as per standard aquaculture procedures.
		c	Administer various stressors to fish in two tanks (i.e., the treatment fish) at selected intervals during rearing and prior to sexual maturation. The stressors should be varied to minimize fish becoming accustomed to one type of stressor.
		d	Spawn treatment and control fish using a single sperm sample and standard aquacultural methods. Record female weight, total length, weight of the eggs, and ovarian fluid weight.
		e	Using subsamples of eggs from each female, weigh to the nearest 0.01 g and measure the diameter to the nearest 0.01 mm each egg in the subsample. Determine the gonadosomatic index, absolute and relative fecundity, and the percent of fertilization.

		f	Determine the mean percent of embryos hatching and monitor growth of fry at 2, 4, 6, and 8 weeks after absorption of the yolk sac. Monitor embryo, fry, and juvenile mortality twice a week.
5	Develop and implement strategies to sustain healthy, viable population.	a	Using information collected formulate risk plan to address future management direction.

Objective schedules and costs

Obj #	Start date mm/yyyy	End date mm/yyyy	Measureable biological objective(s)	Milestone	FY2000 Cost %
1	10/1999	6/2001	Identification of population viability under current fishing regulations	Completion of laboratory experiments	1.00
2	10/2000	6/2003	Identification of population viability under current fishing regulations	Completion of field work	
3	10/2000	10/2003	Identification of population viability under current fishing regulations	Completion of field work	
4	10/2003	9/2004	Identification of population viability under current fishing regulations	Completion of spawning and rearing trials	
5	10/2004	9/2005	Development of risk strategies to protect and perpetuate white sturgeon populations	Altering fisheries management strategies if necessary	
				Total	100.00%

Schedule constraints

Completion date
2004

Section 5. Budget

FY99 project budget (BPA obligated):

FY2000 budget by line item

Item	Note	% of total	FY2000
Personnel	IDFG - \$22,440; USGS - \$82,118	% 39	104,558
Fringe benefits	IDFG - \$8,200; USGS - \$23,814	% 12	32,014
Supplies, materials, non-expendable property	IDFG - \$1,500; USGS - \$44,000	% 17	45,500
Operations & maintenance	IDFG - \$3,000; USGS - \$7,000	% 4	10,000
Capital acquisitions or improvements (e.g. land, buildings, major equip.)		% 0	
NEPA costs		% 0	
Construction-related support		% 0	
PIT tags	# of tags:	% 0	
Travel	IDFG - \$2,000; USGS - \$1,000	% 1	3,000
Indirect costs	IDFG - \$11,400; USGS - \$60,014	% 26	71,414
Subcontractor	University of Idaho	% 2	5,000
Other		% 0	
TOTAL BPA FY2000 BUDGET REQUEST			\$271,486

Cost sharing

Organization	Item or service provided	% total project cost (incl. BPA)	Amount (\$)
IDFG	Field assistance, supervision	% 6	17,700
USGS	Field station support	% 3	8,300
		% 0	
		% 0	
Total project cost (including BPA portion)			\$297,486

Outyear costs

	FY2001	FY02	FY03	FY04
Total budget	\$380,000	\$350,000	\$200,000	\$50,000

Section 6. References

Watershed?	Reference
<input type="checkbox"/>	Barton, B.A., and C.B. Schreck. 1987. Metabolic cost of acute physical stress in juvenile steelhead. Transactions of the American Fisheries Society 116:257-263.
<input type="checkbox"/>	Beggs, G.L., G.F. Holeton, and E.J. Crossman. 1980. Some physiological consequences of angling stress in muskellunge, <i>Esox masquinogoy</i> Mitchell. Journal of Fish Biology.
<input type="checkbox"/>	Biological Risk Assessment Team. 1997. Upper Snake river white sturgeon biological risk assessment. Report for The Nez Perce Tribe.
<input type="checkbox"/>	Birstein, V. J. 1993. Sturgeons and paddlefishes: threatened fishes in need of conservation. Conservation Biology 7(4):773-787.
<input type="checkbox"/>	Booth, R. K., R. S. McKinley, F. Økland, and M. M. Sisak. 1997. In situ measurement of swimming performance of wild Atlantic salmon (<i>Salmo salar</i>) using radio transmitted electromyogram signals. Aquatic Living Resources 10:213-219.
<input type="checkbox"/>	Booth, R. K., J. D. Kieffer, K. Davidson, A. T. Bielak, and B. L. Tufts. 1995. Effects of late-season catch and release angling on anaerobic metabolism, acid-base status, survival, and gamete viability in wild Atlantic salmon (<i>Salmo salar</i>). Canadian Journ
<input type="checkbox"/>	Bouck, G. R., and R. C. Ball. 1966. Influence of capture methods on blood characteristics and mortality in the rainbow trout (<i>Salmo gairdneri</i>). Transactions of the American Fisheries Society 95(2):170-176.
<input type="checkbox"/>	Briggs, C. T., and J. R. Post. 1997. In situ activity metabolism of rainbow trout (<i>Oncorhynchus mykiss</i>): estimates of obtained from telemetry of axial muscle electromyograms. Canadian Journal of Fisheries and Aquatic Sciences 54:859-866.
<input type="checkbox"/>	Campbell, P.M., T.G. Pottinger, and J.P. Sumpter. 1992. Stress reduces the quality of gametes produced by rainbow trout. Biology of Reproduction 47:1140-1150.
<input type="checkbox"/>	Carragher, J.A., J.P. Sumpter, T.G. Pottinger, and A.D. Pickering. 1989. The deleterious effects of cortisol implantation on reproductive function in two species of trout, <i>Salmo trutta</i> L. and <i>Salmo gairdneri</i> Richardson. General and Comparative Endocrinolo
<input type="checkbox"/>	Carragher, J.A., and J.P. Sumpter. 1990. The effect of cortisol on the secretion of sex steroids from cultured ovarian follicles of rainbow trout. General and Comparative Endocrinology 77:403-407.
<input type="checkbox"/>	Carragher, J.A., and N.W. Pankhurst. 1991. Stress and reproduction in a commercially important marine fish, <i>Pagrus auratus</i> (Sparidae). Pages 253-255 in A.P. Scott, J.P. Sumpter, D.E. Kime, and M. Rolfe, editors. Reproductive physiology of fish 1991.
<input type="checkbox"/>	Chan, D.O., and N.Y.S. Woo. 1978. Effect of cortisol on the metabolism of the eel, <i>Anguilla japonica</i> . General and Comparative Endocrinology 35:205-215.

<input type="checkbox"/>	Contreras-Sánchez, W. M. 1995. Effects of stress on the reproductive performance and physiology of rainbow trout (<i>Oncorhynchus mykiss</i>). Master's thesis. Oregon State University, Corvallis, Oregon.
<input type="checkbox"/>	Craig, J. A., and R.L. Hacker. 1940. The history and development of the fisheries of the Columbia River. U.S. Bureau of Fisheries Bulletin 49(32):132-216.
<input type="checkbox"/>	Demers, E., R. S. McKinley, A. H. Weatherly, and D. J. McQueen. 1996. Activity patterns of largemouth and smallmouth bass determined with electromyogram biotelemetry. Transactions of the American Fisheries Society 125:434-439.
<input type="checkbox"/>	DeVore, J. D., B. W. James, C. A. Tracy, and D. A. Hale. 1995. Dynamics and potential production of white sturgeon in the unimpounded lower Columbia river. Transactions of the American Fisheries Society 124:845-856.
<input type="checkbox"/>	Ferguson, R. A., and B. L. Tufts. 1992. Physiological effects of brief air exposure in exhaustively exercised rainbow trout (<i>Oncorhynchus mykiss</i>): implications for catch and release fisheries. Canadian Journal of Fisheries and Aquatic Sciences 49:1157-1
<input type="checkbox"/>	Fielder, D.G., and B.A. Johnson. 1994. Walleye mortality during live-release tournaments on Lake Oahe, South Dakota. North American Journal of Fisheries Management 14:776-780.
<input type="checkbox"/>	Gustaveson, A.W., R.S. Wydoski, and G.A. Wedemeyer. 1991. Physiological response of largemouth bass to angling stress. Transactions of the American Fisheries Society 120:629-636.
<input type="checkbox"/>	Hinch, S. G., R. E. Diewert, T. J. Lissimore, A. M. Prince, M. C. Healey, and M. A. Henderson. 1996. Use of electromyogram telemetry to assess difficult passage areas for river-migrating adult sockeye salmon. Transactions of the American Fisheries Society
<input type="checkbox"/>	Hoefs, N. 1998. Evaluate potential means rebuilding sturgeon populations in the Snake River between Lower Granite and Hells Canyon dams. Phase Two:Multi-Year Study Plan. Prepared for BPA, Contract No. 97AM30423. Nez Perce Tribe, Orofino, ID.
<input type="checkbox"/>	Huuskonen, H., and J. Karjalainen. 1997. Predator-induced respiratory responses in juveniles of vendace <i>Coregonus albula</i> , whitefish <i>C. lavaretus</i> , perch <i>Perca fluviatilis</i> and roach <i>Rutilus rutilus</i> . Environmental Biology of Fishes 49:265-269.
<input type="checkbox"/>	Kaseloo, P.A., A.H. Weatherly, J. Lotimer, and M.D. Farina. 1992. A biotelemetry system recording fish activity. Journal of Fish Biology 40:165-179.
<input type="checkbox"/>	Laitinen, M., and T. Valtonen. 1994. Cardiovascular, ventilatory and total activity responses of brown trout to handling stress. Journal of Fish Biology 45:933-942.
<input type="checkbox"/>	Lucas, M. C., I. G. Priede, J. D. Armstrong, A. N. Z. Gindy, and L. De Vera. 1991. Direct measurements of metabolism, activity and feeding behaviour of pike, <i>Esox lucius</i> L., in the wild, by the use of heart rate telemetry. Journal of Fish Biology 39:325-34
<input type="checkbox"/>	Lucas, M.C., A.D.F. Johnstone, and I.G. Priede. 1993. Use of physiological

	telemetry as a method of estimating metabolism of fish in the natural environment. Transactions of the American Fisheries Society 122:822-833.
<input type="checkbox"/>	Mazeaud, M. M., M. Frederic, and E. M. Donaldson. 1977. Primary and secondary effects of stress in fish: some new data with a general review. Transactions of the American Fisheries Society. 106(3):201-212.
<input type="checkbox"/>	McKinley, R.S. and G. Power. 1992. Measurement of activity and oxygen consumption for adult lake sturgeon (<i>Acipenser fulvescens</i>) in the wild using radio-transmitter EMG signals. Pages 307-318 in I.G. Preide and S.M. Swift, eds. Wildlife telemetry: remote
<input type="checkbox"/>	Melotti, P., A. Roncarati, E. Garella, O. Carnevali, G. Mosconi and A. Polzonetti-Magni. 1992. Effects of handling and capture stress on plasma glucose, cortisol and androgen levels in brown trout, <i>Salmo trutta morpha fario</i> . Journal of Applied Ichthyology
<input type="checkbox"/>	Miller, A. I., T. D. Counihan, M. J. Parsley, and L. G. Beckman. 1995. Columbia river basin white sturgeon. Pages 154-157 in LaRoe, E.T., G.S. Farris, C.E. Puckett, P.D. Doran, and M.J. Mac, editors. Our Living Resources: a report to the nation on the dis
<input type="checkbox"/>	Morgan, J.D., and G.K. Iwama. 1996. Cortisol-induced changes in oxygen consumption and ionic regulation in coastal cutthroat trout (<i>Oncorhynchus clarki clarki</i>) parr. Fish Physiology and Biochemistry 15(5):385-394.
<input type="checkbox"/>	Pankhurst, N. W., and M. Dedual. 1994. Effects of capture and recovery on plasma levels of cortisol, lactate and gonadal steroids in a natural population of rainbow trout. Journal of Fish Biology 45:1013-1025.
<input type="checkbox"/>	Peters, G., M. Faisal, T. Lang, and I. Ahmed. 1988. Stress caused by social interaction and its effect on susceptibility to <i>Aeromonas hydrophila</i> infection in rainbow trout <i>Salmo gairdneri</i> . Diseases of Aquatic Organisms 4:83-89.
<input type="checkbox"/>	Pickering, A.D., T.G. Pottinger, J. Carragher, and J.P. Sumpter. 1987. The effects of acute and chronic stress on the levels of reproductive hormones in the plasma of mature male brown trout, <i>Salmo trutta</i> L. General and Comparative Endocrinology 68:249-25
<input type="checkbox"/>	Priede, I. G., and P. Tytler. 1977. Heart rate as a measure of metabolic rate in teleost fishes; <i>Salmo gairdneri</i> , <i>Salmo trutta</i> and <i>Gadus morhua</i> . Journal of Fish Biology 10:231-242.
<input type="checkbox"/>	Priede, I.G. and A.H. Young. 1977. The ultrasonic telemetry of cardiac rhythms of wild brown trout (<i>Salmo trutta</i> L.) as an indicator of bio-energetics and behaviour. Journal of Fish Biology 10:299-318.
<input type="checkbox"/>	Rogers, S.C., and A.H. Weatherly. 1983. The use of opercular muscle electromyograms as an indicator of the metabolic costs of fish activity in rainbow trout, <i>Salmo gairdneri</i> Richardson, as determined by radiotelemetry. Journal of Fish Biology 23: 535-547.
<input type="checkbox"/>	Schreck, C.B. and P.B. Moyle, editors. 1990. Methods for fish biology. American Fisheries Society, Bethesda, Maryland.
<input type="checkbox"/>	Sumpter, J.P., J.F. Carragher, T.G. Pottinger, and A.D. Pickering. 1987. Interaction of stress and reproduction in trout. Pages 299-302 in D.R. Idler, L.W. Crim, and J.M. Walsh, editors. Reproductive physiology of fish 1987. St. Johns: Memorial University

<input type="checkbox"/>	Sureau, D., and J. P. Lagardère. 1991. Coupling of heart rate and locomotor activity in sole, <i>Solea solea</i> (L.), and bass, <i>Dicentrarchus labrax</i> (L.), in their natural environment by using ultrasonic telemetry. <i>Journal of Fish Biology</i> 38:399-405.
<input type="checkbox"/>	Tomasso, A. O., J. J. Isely, and J. R. Tomasso, Jr. 1996. Physiological responses and mortality of striped bass angled in freshwater. <i>Transactions of the American Fisheries Society</i> 125:321-325.
<input type="checkbox"/>	Tufts, B.L., Y. Tang, K. Tufts, and R. G. Boutilier. 1991. Exhaustive exercise in ? wild? Atlantic salmon (<i>Salmo salar</i>): acid-base regulation and blood gas transport. <i>Canadian Journal of Fisheries and Aquatic Sciences</i> 48:868-874.
<input type="checkbox"/>	Weatherly, A.H., S.C. Rogers, D.G. Pincock, and J.R. Patch. 1982. Oxygen consumption of active trout, <i>Salmo gairdneri</i> R., derived from electromyograms obtained from radiotelemetry. <i>Journal of Fish Biology</i> 20:479-489.
<input type="checkbox"/>	Wilkie, M.P., and six coauthors. 1996. Physiology and survival of wild Atlantic salmon following angling in warm summer waters. <i>Transactions of the American Fisheries Society</i> 125:572-580.
<input type="checkbox"/>	Wood, C.M., J.D. Turner, and M.S. Graham. 1983. Why do fish die after severe exercise? <i>Journal of Fish Biology</i> 22:189-201.
<input type="checkbox"/>	Wydoski, R. S., G. A. Wedemeyer, and N. C. Nelson. 1976. Physiological response to hooking stress in hatchery and wild rainbow trout (<i>Salmo gairdneri</i>). <i>Transactions of the American Fisheries Society</i> 5:601-606.

PART II - NARRATIVE

Section 7. Abstract

The white sturgeon is a species critically affected by hydroelectric development. Because of the declining status of many sturgeon populations, catch and release angling has become a central part of current fisheries management activities directed at sustaining or recovering populations of these fish. However, the potential detrimental effects of catch and release angling on these fish have never been addressed. We propose to examine the effects of catch and release angling on the stress physiology, reproductive physiology, and mortality of white sturgeon in laboratory and field studies. This research will use a multi-disciplinary approach that will focus on state-of-the-science physiological telemetry techniques to assess the effects of angling stress on fish in the wild. Data from the laboratory on the physiological responses and metabolic costs of physical stress in white sturgeon will be coupled with metabolic rate data obtained by physiological telemetry from fish angled in the wild to allow an assessment of catch and release angling. This project should be relevant to the resident fish goal and sturgeon mitigation as described in section 10 of the FWP. Results from this study should be complete within 3-4 years and will enable fisheries managers to make more informed decisions regarding the use of catch and release angling as a management tool for sustaining or recovering populations of white sturgeons. The study could be used to decide if seasonal closures to

angling are warranted to protect spawning fish or to improve survival of fish during the summer when water temperatures are at their highest.

Section 8. Project description

a. Technical and/or scientific background

The white sturgeon is one of the Pacific Northwest's most important recreational fish and currently supports the largest sport fishery in terms of effort in the Columbia Basin (Devore et al. 1995). However, several populations of white sturgeon in the Columbia Basin are sparse and declining, primarily as a result of hydroelectric development. This has resulted in a variety of management activities by state agencies to help preserve sensitive stocks, including angling regulations such as slot-length limits to protect sexually mature fish, reduced harvest limits and seasons, and the use of barbless hooks only. Regulations such as these are necessary since white sturgeon in the Columbia Basin have a long history of stock collapse due to overexploitation (Craig and Hacker 1940). In fact, the longevity, slow growth, and delayed maturation of sturgeons make them particularly vulnerable to overexploitation and changes in their environment (Birstein 1993; DeVore et al. 1995; Miller et al. 1995).

Because of current angling regulations and increased recognition by anglers that trophy-sized fish can be readily hooked, increasing numbers of white sturgeon are now being released shortly after they are landed. Opportunities to hook and land trophy white sturgeons are promoted by a number of commercial fishing guides throughout the basin. Although catch and release angling is widespread and is actively promoted as a method to conserve white sturgeon stocks, nothing is known about the biological effects of catch and release angling for this species. Information on the post-angling survival and physiology of white sturgeon would be helpful to fishery managers in evaluating current and developing new regulations to sustain these sensitive fish stocks.

Angling is one of the most severe forms of exhaustive exercise that a fish can experience (Booth et al. 1995). Many studies have shown that exhaustive exercise, including angling, results in a variety of severe physiological disturbances in different species of fish (Wydoski et al. 1976; Wood et al. 1983; Tufts et al. 1991; Pankhurst and Dedual 1994; Booth et al. 1995; Tomasso et al. 1996; Wilkie et al. 1996). Several studies have also documented delayed mortality after exhaustive exercise or angling for a variety of species, including rainbow trout *Oncorhynchus mykiss* (Bouck and Ball 1966; Ferguson and Tufts 1992), Atlantic salmon *Salmo salar* (Wilkie et al. 1996), muskellunge *Esox masquinongy* (Beggs et al. 1980), walleye *Stizostedion vitreum* (Fielder and Johnson 1994), and striped bass *Morone saxitalis* (Tomasso et al. 1996). In contrast, other investigators have found little association between exhaustive exercise and post exercise mortality (Wydoski et al. 1976; Gustaveson et al. 1991; Tufts et al. 1991; Pankhurst and Dedual 1994). Discrepancies between studies not only indicate that additional factors may influence postangling survival (Wilkie et al. 1996) but also presents difficulties in attempting to apply these results to unstudied species.

In addition to physiological dysfunction and delayed mortality, exhaustive exercise (and many other types of stressors) may alter several aspects of reproduction in fish. For example, several studies have demonstrated that stress may severely alter levels

of reproductive hormones in fish (Pickering et al. 1987; Carragher et al. 1989; Carragher and Pankhurst 1991; Melotti et al. 1992; Pankhurst and Dedual 1994). Cortisol, the corticosteroid hormone secreted by fish in response to a variety of environmental stressors, has been shown to inhibit production of estradiol and testosterone by ovarian follicles of rainbow and brown *S. trutta* trout (Sumpter et al. 1987; Carragher and Sumpter 1990). Campbell et al. (1992) reported that exposure of female hatchery rainbow trout to acute stress resulted in smaller egg size, delayed ovulation, and lower survival of larvae compared to unstressed fish. In contrast, Booth et al. (1995) found no significant differences in the hatching success of eggs between groups of angled and nonangled Atlantic salmon, but cautioned that angling may affect other aspects of reproduction, such as spawning behavior.

Although some areas of the Columbia River have been closed to angling prior to and during the spawning period for white sturgeon, many areas remain open to angling during this time. Also, the size limit regulations for white sturgeon that are in effect do protect sexually mature fish from being harvested, but such fish are nevertheless commonly captured and then released for sport. All white sturgeon in the Snake River in Idaho are under catch and release regulations year around. Clearly, an understanding of the effects of catch and release angling on reproductive physiology is necessary for a complete evaluation of this practice.

Physiological telemetry has proven to be a useful tool for evaluating various aspects of the physiology and behavior of fish in the wild. This technique involves recording transmissions of physiological parameters such as heart rate, ventilation rate, or axial muscle contractions that are strongly correlated with oxygen consumption. For example, detailed *in situ* measurements of locomotor activity, selected aspects of metabolism, and feeding activity have been made for a variety of fishes using transmissions of heart rate (Priede and Tytler 1977; Lucas et al. 1991; Sureau and Lagardere 1991), ventilation rate (Rogers and Weatherly 1983), and axial muscle electromyograms (EMGs; Weatherly et al. 1982; Demers et al. 1996; Hinch et al. 1996; Booth et al. 1997; Briggs and Post 1997).

Because of the success of physiological telemetry in evaluating the physiology and behavior of fish in the wild, we believe this technique will allow a rigorous assessment of the stressful effects of catch and release angling on white sturgeon. Stress in fish is also known to increase heart rate, ventilation rate, and oxygen consumption (Mazeaud et al. 1977; Barton and Schreck 1987; Peters et al. 1988; Laitinen and Valtonen 1994; Huuskonen and Karjalainen 1997). For example, Laitinen and Valtonen (1994), using a biotelemetry system, found that both heart and ventilation rate of brown trout were elevated for 3 to 4 days following a handling stress. Barton and Schreck (1987) documented that minor physical disturbances elicited a more than twofold increase in the metabolic rate of juvenile steelhead and estimated that the energetic cost of such stress was about one-quarter of the scope for activity of the fish. In addition, there is increasing evidence of a positive relation between plasma cortisol level and metabolic rate (i.e., oxygen consumption) in stressed fish (Chan and Woo 1978; Barton and Schreck 1987; Morgan and Iwama 1996) which we believe substantiates the use of metabolic rate as an indicator of stress in fish. By combining laboratory derived relations between metabolic rate and physiological variables (i.e., heart rate, ventilation rate, EMGs, and classic

physiological indicators of stress) with telemetered data from white sturgeon angled in the wild, it should be possible to provide a unique and powerful means of assessing the energetic costs of capture and release, the time needed for recovery, and the extent of any post-angling mortality.

b. Rationale and significance to Regional Programs

Current fisheries management activities directed at sustaining or recovering populations of white sturgeon are a direct response to the deleterious effects of hydroelectric development. We consider catch and release angling as a management strategy responding to the detrimental effects of the hydroelectric system and therefore believe this strategy requires a complete evaluation to help assess its efficacy. Already, angling for white sturgeons has been prohibited in two areas--the Kootenai River in northern Idaho, where white sturgeons are listed as endangered, and that portion of the Columbia River upstream from the international border between the U.S. and Canada. These areas were closed to angling because of concerns that the stress associated with handling could either cause mortality or reproductive failure in mature fish. Fisheries managers recognize the value in providing the public with the opportunity to fish for and potentially catch large white sturgeons. However, the effects of catch and release fishing on the survival or reproduction of these fish have never been addressed.

The recently completed Biological Risk Assessment for the Upper Snake River White Sturgeon (Biological Risk Assessment Team 1997), prepared for the Nez Perce Tribe under section 10.4A.4 of the FWP and funded by the BPA, listed the need to evaluate the consequences of catch and release fishing as the *highest* research priority to address in the efforts to recover the population of white sturgeons found in the Snake River between Lower Granite Dam and Hells Canyon Dam. The project described herein is relevant to the resident fish goal as outlined in section 10.1 of the FWP in that it will provide much needed information to help assess measures designed to protect and recover white sturgeon stocks. This project specifically addresses several priorities for implementing resident fish policies and projects, as discussed in section 10.1B of the 1995 amendments to the FWP, including “accord highest priority to weak, but recoverable, native populations injured by the hydropower system..”and “accord high priority to populations that support important fisheries...including sturgeon...”. Finally, this project is clearly relevant to several aspects of section 10.4 of the FWP dealing with sturgeon mitigation. This project can be completed relatively quickly, has a high potential for success and learning new information, addresses concerns being discussed by the Nez Perce tribe (as described above), and may be helpful to other agencies or tribes in addressing the extent of fishing mortality in white sturgeon (e.g., section 10.4A.6 of the 1995 amendments to the FWP).

c. Relationships to other projects

We're not aware of an Umbrella Project for white sturgeon projects. However, there is a high level of coordination among researchers and managers in the basin who work with this species. The biological objectives and management plans for white sturgeon populations in the basin all call for sustainable levels of recreational and subsistence

harvest. It's recognized that when fishing is allowed, there will always be catch and release fishing on sublegal and over-legal sized fish. This proposed project will address concerns that fishery managers may have regarding this activity.

Projects 860500, *White Sturgeon Mitigation and Restoration in the Columbia and Snake Rivers*, 9700900, *Evaluate Means of Rebuilding White Sturgeon Populations in the Lower Snake River*, 9093, *Consumptive Sturgeon Fishery – Hells Canyon and Oxbow Reservoirs*, and 9502700, *Assess Limiting Factors of the Lake Roosevelt White Sturgeon Population*, have management goals of providing increased fishing opportunities. This proposed project will complement those efforts.

Equipment and expertise from Project 860500 will be used extensively on this project. Significant cost savings will be realized in capturing fish and conducting the telemetry work necessary to complete this work.

The fourth objective of this proposal will require the use of an aquaculture facility for holding, stressing, and subsequently spawning and rearing white sturgeons. We will coordinate with several ongoing projects including 860500, 8806400, *Kootenai River White Sturgeon Studies and Conservation Aquaculture*, and 9603201, *Begin Implementation of Year 1 of the K-Pool Master Plan Program*, to identify locations to conduct this work.

d. Project history (for ongoing projects)

This is a new project.

e. Proposal objectives

1. Examine the relation between telemetered physiological variables, oxygen consumption, and physiological indicators of stress in white sturgeon.

Ho: There is no significant relation between physiological telemetry (i.e., heart rate, ventilation rate, or EMG) output and oxygen consumption.

Ho: There is no significant relation between oxygen consumption and physiological indicators of stress.

2. Assess the effects of catch and release angling on metabolic rate and selected physiological indicators of stress in wild white sturgeon.

Ho: Catch and release angling is not stressful to white sturgeon.

3. Document the occurrence and determine the extent of any postangling mortality.

Ho: Catch and release angling does not cause significant postangling, delayed mortality in white sturgeons.

4. Assess the effects of exhaustive stress on selected aspects of white sturgeon reproductive performance.

Ho: Catch and release angling has no significant effects on reproduction of individuals in white sturgeons.

f. Methods

Objective 1. Examine the relation between telemetered physiological variables, oxygen consumption, and physiological indicators of stress in white sturgeon. We will use swimming respirometry to determine the relation between metabolic rate (i.e., oxygen consumption) and the physiological variable of choice (i.e., heart rate, ventilation rate, or EMG's; see Lucas et al. (1993) for a complete review of methodology). We will obtain or build a Blazka or Brett-type swimming respirometer large enough to swim sturgeon from about 0.9-1.5 m in length. We will obtain fish from the wild by angling or set-lining and transport them to our laboratory. Fish will be held in large circular tanks at the CRR laboratory under low densities and ambient environmental conditions. They will be fed daily with live or dead juvenile salmonids. Fish will be held for at least two weeks prior to the start of respirometry trials. We anticipate needing only a few (e.g., 6-10) fish to complete this objective.

We will obtain a few sonic or radio tags designed to telemeter either heart rate, ventilation rate, or EMG's from established manufacturers. We will also obtain all equipment necessary to monitor telemetered signals and determine the ideal operating conditions for the system. Much of this equipment we already have at CRR laboratory. Tags will be surgically implanted in fish according to methods described in Priede and Young (1977), Kaseloo et al. (1992), and McKinley and Power (1992). In addition to the these references, we will obtain hands-on instruction in surgical tag implantation from Steve Peake at Simon Fraser University, who has extensive experience with the techniques involved. We will also surgically implant cannulas into fish for repeated sampling of blood during respirometry trials. Cannulas will be placed in either the dorsal aorta, branchial vasculature, or caudal vasculature according to standard techniques (see Schreck and Moyle 1990). Surgeries will be performed on all fish during a 1-3 day period, after which fish will be held in their tanks for at least a week to recover and to test the telemetry system.

To begin a respirometry trial, a fish will be chosen at random, placed in the respirometer, and allowed a period of acclimation to low water velocity. During acclimation, the swimming chamber will be flushed with oxygen-saturated freshwater. There will likely be an electric grid at the downstream end of the swim chamber to encourage fish to constantly swim. After acclimation, we will subject each fish to a U-critical swimming test by incrementally increasing water velocity at set time intervals until fish become fatigued. Oxygen concentrations in the respirometer will be determined at set intervals (e.g., every 5 min) using calibrated oxygen meters or Winkler titrations of water samples. Telemetered data will be continuously recorded and averaged over the set intervals. Blood samples will also be taken periodically during and after the swimming trial. Experiments will be carried out over the range of temperatures likely to be experienced by these fish in the wild.

Resultant data will be analyzed using regression analysis and analysis of covariance to determine the nature of the relation between oxygen consumption (i.e., metabolic rate) and the physiological variable of interest. Based on the literature, we fully expect at least one physiological variable to be highly correlated with metabolic rate.

Blood samples will be assayed for various indicators of stress, including but not limited to such factors as plasma cortisol and lactate. We will also assess the relation between such classical indicators of stress and metabolic rate.

Objective 2. Assess the effects of catch and release angling on metabolic rate and selected physiological indicators of stress in wild white sturgeon. Based on our laboratory results from the first objective, we will obtain several (e.g., 10-15) physiological sonic or radio tags that will telemeter the variable of choice. We will capture sturgeon from the Snake River using angling or set lines and either surgically implant tags *in situ* or transport fish to a suitable site for tag implantation. Our preference is to implant tags *in situ*, but our results from objective 1 may indicate that implantation under more controlled conditions is more prudent. In addition, it may be desirable to double tag fish with two different types of tags. For example, EMG tags may provide information during the hooking and playing process, whereas heart rate tags may yield the best metabolic rate information after release. After tag implantation, fish will be released back to their point of capture.

After release, USGS and IDFG personnel will monitor physiological function and fish movement for several weeks to establish baseline metabolic rate data. Data should be collected for a period sufficient to account for recovery from surgery and diel variations in metabolic rate. After collecting sufficient baseline metabolic rate data, we will use the telemetry system to precisely locate the fish and attempt to capture the tagged fish by angling. This technique has been used successfully in other areas. For this concentrated, intense angling effort, we will use one or both of these strategies: (1) agency boats equipped with several rod and reel outfits or (2) enlist private sportsmen or angling groups to conduct the angling. We surmise this project would be of interest to anglers and to maximize participation could offer a monetary prize to the angler catching the tagged fish. When a tagged fish is hooked, which should be indicated by an abrupt change in tag output, we will note the time of hookup and the playing time. We will monitor physiological function during the angling process using tag output. Upon landing the fish, we will draw a quick blood sample, photograph the fish and the site of surgery, and release the fish.

After release, we will monitor physiological function of fish for several days in a manner identical to that during baseline data collection. Telemetered physiological data, in conjunction with our laboratory-derived relations, will be used to estimate oxygen consumption of angled fish. We will then compare oxygen consumption of angled fish at various intervals post-release to that of resting laboratory fish and the baseline data to determine the metabolic cost (i.e., the stressful effects) of catch and release angling. Plasma samples taken from fish in the wild will be assayed for physiological indicators of stress and values compared with those from fish used in the respirometry experiments. Because of the data intensive nature of this work and other logistical considerations, we anticipate tagging and releasing less than 10 fish per field season.

Objective 3. Document the occurrence and determine the extent of any postangling mortality. Work addressing this objective will be conducted simultaneously with that of objective 2. Basically, the USGS and IDFG will monitor the physiology and movements of fish post-release for a period of time sufficient to allow a reasonable determination of

whether the fish survived. We will use a lack of tag output and fish movements as evidence, but not conclusive proof, that an angled fish has not survived. There are two critical assumptions associated with this objective: (1) the fish has not lost its tag and (2) the tag is functioning properly during the post-release monitoring period. Two pieces of information will help in meeting these assumptions: (1) when the fish is landed after angling, we will be able to assess the condition of the surgery site and hence obtain some indication of the possibility of losing the tag in the future and (2) the tags should be large enough to have a battery that will last several months. At the end of the field season, we will simply tally the number of fish that may not have survived the angling event.

Objective 4. Assess the effects of various physical stressors on selected aspects of white sturgeon reproductive performance. Work conducted for this objective will be based upon that described by Contreras-Sanchez (1995). We will establish facilities at our laboratory for the long-term rearing of adult sturgeon or, if necessary, will obtain rearing space at an established aquaculture facility. We will obtain several reproductive age (or size) female sturgeon from the wild or from an established aquaculture facility. Fish will be stocked into each of 4 large, circular tanks that will receive water of ambient temperature during the year. Fish will be fed a diet of live and dead juvenile salmonids. There will be two treatments, stressed and controls, with two replicate tanks per treatment.

During rearing, we will monitor reproductive physiology and state of maturation by periodically sampling blood for sex steroid analysis and conducting ovarian biopsies using standard aquacultural procedures. These sampling episodes will be at the minimum necessary to increase our chances of successful spawning. We will also administer various stressors to fish in two tanks (i.e., the treatment fish) at selected intervals during rearing and prior to sexual maturation. We will vary the stressors to minimize fish becoming accustomed to one type of stressor. The stressors may include, but will not be limited to, dewatering and hypoxia, chasing the fish to exhaustion, and netting fish and suspending in air.

When fish are sexually mature, we will spawn all treatment and control fish using standard aquacultural procedures. We will use a single, pooled sperm sample from several males to fertilize the eggs. We will record female weight, total length, weight of the eggs, and ovarian fluid weight. All eggs will be incubated in separate chambers. Using subsamples of eggs from each female, we will weigh and measure the diameter of each egg in the subsample (ca. 50-100 eggs per female). The gonadosomatic index, absolute and relative fecundity, and the percentage of fertilized eggs for each female will be determined. We will determine the mean percent of embryos hatching and monitor growth of fry at 2, 4, 6, and 8 weeks after absorption of the yolk sac. We will monitor and tally embryo, fry, and juvenile mortality twice a week.

We will use analysis of variance (ANOVA) as a check on randomization for a similar distribution of fish size throughout all tanks. Data from within each tank will be pooled for analysis, and we will check for tank effects between replicates using t-tests, or, if data distribution warrants it, their nonparametric equivalents. For all pairwise comparisons between treatments, we will use t-tests or nonparametric equivalents. We will use correlation and regression analyses to assess the relation between egg and fry size, and egg size and mortality and compare the relations between treatments. The level

of significance for all tests will be 0.05.

Objective 5. Develop and implement strategies to sustain healthy, viable population.

The ultimate goal of this proposal will be to address impacts on white sturgeon populations as a result of sport angling. Under this objective, we will use the results obtained from completing the previous objectives to assist in setting sport fishing regulations.

g. Facilities and equipment

We anticipate that all of the laboratory work for this project will be conducted at our Columbia River Research facility. Our laboratory, which has a long history of conducting research throughout the basin, has a fully equipped wet laboratory as well as several dry laboratories for conducting physiological assays. Our hope is to borrow a respirometer for completion of objective 1, but one could be constructed if necessary. We plan on drawing on the experience of Dr. Joseph Cech of UC-Davis to assist in this endeavor. Our staff has several individuals who are well versed in the art and science of wildlife telemetry, and we plan on using their experience to assist us in this project. In addition, we have an extensive array of state-of-the-art telemetry equipment from which to draw upon to conduct this research. Our laboratory is also well equipped with a variety of vessels of all sizes to conduct a wide array of field work. Our office is well supplied with all the modern equipment, computers, and analysis software necessary to complete this research. In short, our laboratory already has much of the equipment and technology necessary to complete this research, which we believe will result in substantially lower costs.

The field exercise to monitor white sturgeon during sport angling will be conducted in the Snake River near Lewiston, ID. The IDFG Clearwater Region office is located in Idaho and will support that portion of the project. IDFG has suitable boats and other equipment to support the project. In addition, other necessary equipment (computers, etc.) are available for use at that office.

h. Budget

Personnel costs include partial salaries for the project co-managers and salaries for research and technical staff to conduct the experiments. Supplies include the construction of a respirometer large enough to swim mature white sturgeons, miscellaneous laboratory supplies and physiological transmitters. Operations and Maintenance costs include boat and vehicle costs for capturing and transporting fish, and holding fish in the wet lab. The USGS assessment rate is set at 38% of all direct costs.

The University of Idaho will provide student assistance through a subcontract.

Section 9. Key personnel

Matthew G. Mesa, Research Fishery Biologist, 1.0 FTE

Experience

- 1991-Present Research Fishery Biologist, U.S. Geological Survey, Biological Resources Division, Columbia River Research Lab, Cook, WA
Current responsibilities: Team leader on research projects addressing the effects of thermal stress on juvenile salmonids and evaluating the swimming performance and physiology of Pacific lamprey
- 1989-1991 Fishery Biologist, U.S. Fish and Wildlife Service, Seattle-NFRC, Columbia River Field Station, Cook, WA
- 1986-1989 Fishery Biologist/CEA Appointee, Seattle-NFRC, Oregon Cooperative Fisheries Research Unit, Oregon State University, Corvallis, OR
- 1984-1986 Fishery Biologist, U.S. Fish and Wildlife Service, Seattle-NFRC, Columbia River Field Station, Cook, WA

<u>Education:</u>	<u>School</u>	<u>Degree and Date Received</u>
	California Polytechnic State Univ. at San Luis Obispo	B.S., Nat. Res. Mgt., 1984
	Oregon State Univ.	M.S., Fisheries Science, 1989
	Oregon State Univ.	Ph.D, 1999 (anticipated)

Expertise: My areas of expertise include predator-prey interactions in fishes, fish behavior and performance, and general and stress physiology of fishes

Publications and Reports

- Mesa, M.G. and C.B. Schreck. 1989. Electrofishing mark-recapture and depletion methodologies evoke behavioral and physiological changes in cutthroat trout. Transactions of the American Fisheries Society 118:644-658.
- Mesa, M.G., and T.M. Olson. 1993. Prolonged swimming performance of northern squawfish. Transactions of the American Fisheries Society 122:1104-1110.
- Mesa, M.G. 1994. Effects of multiple acute stressors on the predator avoidance ability and physiology of juvenile chinook salmon. Transactions of the American Fisheries Society 123:786-793.
- Mesa, M.G., T.P. Poe, D.M. Gadomski, and J.H. Petersen. 1994. Are all prey created equal? A review and synthesis of differential predation on prey in substandard condition. Journal of Fish Biology 45 (Supplement A):81-96.
- Mesa, M.G., T.P. Poe, A.G. Maule, and C.B. Schreck. *In press*. Vulnerability to predation and physiological stress responses in juvenile chinook salmon experimentally infected with *Renibacterium salmoninarum*. Canadian Journal of Fisheries and Aquatic Sciences.

Michael J. Parsley

Research Fishery Biologist - Project Leader 0.10 FTE

School	Degree	Date
Iowa State University	B.S. Fisheries & Wildlife Biology	1982
University of Wisconsin at Stevens Point	M. S. Fisheries	1984

Certified by the American Fisheries Society as a Fisheries Scientist in 1990

Current Employer: U.S. Geological Survey - Biological Resources Division

Current Responsibilities: I serve as project leader for studies done by staff at our facility on the early life history and habitat use of white sturgeons in the Columbia River. The studies have included the use of biotelemetry to ascertain habitat use by juvenile and adult white sturgeons, laboratory experiments to determine the effects of gas supersaturation on developing embryos, and the use of trawls to estimate recruitment to young of the year. My role is to coordinate our research activities on white sturgeons with the activities and needs of the tribes, states, and other governmental agencies. I oversee the work of several biologists and technicians who collect and analyze data pertaining to our studies to ensure that our work is of the highest quality and that it is done in accordance with established standards and protocols, such as the Good Laboratory Practices Act.

I also serve as the geospatial technology coordinator for the Western Fisheries Research Center.

Recent Previous Employment: Research Fisheries Biologist, U.S. Geological Survey, Biological Resources Division, Columbia River Research Laboratory, 1984 - present.

Expertise: My area of expertise is Fisheries Research, and I'm considered an expert on the ecology and biology of white sturgeons. In 1993 I organized and co-chaired a day-long symposium called "Biology and Management of North American Sturgeons" that was held at the Annual Meeting of the American Fisheries Society, Portland, Oregon, 1993. I'm also knowledgeable in methods to quantify habitat in large rivers using remote sensing and geographic information systems.

Selected Relevant Publications:

Parsley, M. J., L. G. Beckman, and G. T. McCabe. 1993. Spawning and rearing habitat use by white sturgeons in the Columbia River downstream from McNary Dam. Transactions of the American Fisheries Society 122:217-227.

Parsley, M. J., and L. G. Beckman. 1994. White sturgeon spawning and rearing habitat in the lower Columbia River. North American Journal of Fisheries Management 14:812-827.

Counihan, T.D., A.I. Miller, M.G. Mesa, and M.J. Parsley. 1998. The effects of dissolved gas supersaturation on white sturgeon larvae. Transactions of the American Fisheries Society. 127:316-322.

Counihan, T.D., A.I. Miller, and M.J. Parsley. In press. Indexing the relative abundance of young-of-the-year white sturgeon in an impoundment of the lower Columbia River from highly skewed trawling data. North American Journal of Fisheries Management.

Tim Cochnauer, PhD 0.1 FTE
Regional manager
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Education: Doctorate in Fishery Resources, 1983, University of Idaho, Moscow, ID
MS in Zoology, 1973, University of Oklahoma, Norman, OK
BS in Zoology, 1967, University of Oklahoma, Norman, OK

Current responsibilities: As regional fish manager, the project manager responsible for both anadromous and resident fish populations and fisheries within the Clearwater Region of north central Idaho. The area encompasses the entire Clearwater River drainage, the Snake River drainage upstream to Hells Canyon Dam, the Palouse River drainage, and the Salmon River drainage (north side) from its mouth upstream to Horse Creek (rkm 300). The IDFG regional fish staff is comprised of four fishery scientists conducting a variety of activities including data collection, creel census, fish management decisions and establishing and implementing sport fishing regulation.

This individual has over 25 years experience with the Idaho Department of Fish and Game working in fish research, fish management and the hatchery system. Experience includes radio-tagging and monitoring a variety of fish species throughout Idaho, including white sturgeon.

Publications:

Cochner, T. 1983. Abundance, distribution, growth and management of White Sturgeon *Acipenser transmontanus* in the Middle Snake River Idaho. Doctoral Dissertation, University of Idaho, Moscow, ID

Cochner, T., J.R. Lukens, F.E. Partridge. 1985. Status of white sturgeon, *Acipenser transmontanus*, in Idaho. Pages 127-133 in F.P. Binkowski and S.I. Doroshov (eds) North American Sturgeons: biology, and aquacultural potential. Dr. W. Junk, Dordrecht, The Netherlands.

Cochner, T., E. Schriever, and J. Brostrom. 1996. River and Stream Investigations. F-71-20. Federal Aid in Sport Fish Restoration. Idaho Department of Fish and Game.

Cochner, T., E. Schriever, and J. Brostrom. 1997. River and Stream Investigations. F-71-21. Federal Aid in Sport Fish Restoration. Idaho Department of Fish and Game.

Cochner, T., E. Schriever, and J. Brostrom. 1998. River and Stream Investigations. F-71-22. Federal Aid in Sport Fish Restoration. Idaho Department of Fish and Game.

Section 10. Information/technology transfer

Research results will be submitted for publication in peer-reviewed journals. The work will also be presented at various meetings and workshops.

Congratulations!